

EFFECT OF ACTIVATION TIME ON THE PINANG FROND BASED ACTIVATED CARBON FOR REMAZOL BRILLIANT BLUE R REMOVAL

M.A. Ahmad*, S.G. Herawan, and A.A. Yusof

Center of Advanced Research on Energy (CARE) Faculty of Mechanical Engineering UniversitiTeknikal Malaysia Melaka, Hang Tuah Jaya, 76100 Durian Tunggal, Melaka, MALAYSIA. *E-mail: azharahmad.utem@gmail.com

ABSTRACT

Activated carbons are regularly used for dye wastewater treatment. They can be produced from various organic materials which have a high level of carbon content. In this study, Pinang frond based activated carbon (PFAC) was produced by using a physical activation process to study the effect of different activation times in the range of 1–7 hours. The optimum PFAC sample was found with an activation time of 3 hours, which gives the highest BET surface area and pore volume of 958.23 m²/g and 0.5469 mL/g, respectively. This sample shows well-developed pore structure with a high fixed carbon content of 79.74%. The removal of 87.6% of Remazol Brilliant Blue R (RBBR) was achieved for a sample with initial RBBR concentration of 50 mg/L, and 40.8% was achieved with 500 mg/L. The results indicated that the PFAC is very effective for RBBR adsorption from aqueous solution.

Keywords: Activated carbon; Pinang frond; activation time; Remazol brilliant blue R

INTRODUCTION

Activated carbon has been proven to be an effective and widely used adsorbent for the removal of a variety of organic and inorganic pollutants dissolved in aqueous solution [1], or from a gaseous environment [2] Activated carbons used as adsorbent have a comparatively high surface area and total pore volume. The commercially available activated carbons made from petroleum coke, bituminous coal, and lignite are expensive, which limits their use and application [3]. Therefore, other alternative precursors which are inexpensive and abundantly available for activated carbon production have been investigated. From the literature, oil palm stone [4], pinang frond [5], banana stalk [6], bamboo species [7], coffee husk [8], date pits [9], oil palm shell [10] and Parkinsoniaaculeatea wood [11] have all been used to prepare activated carbon.

This activated carbon can be produced using physical and chemical activation treatments. The advantages of physical activation are the possibility of developing a structure with more pores and large active area [9] and having less effect in terms of the secondary pollution problem during the disposal stage [12]. The most important characteristic of an activated carbon is its adsorption capacity or uptake, which is highly influenced by the preparation conditions of activated carbon, such as activation time. The activation time is able to influence the activated carbon prepared, as prolonged activation time entails the opening and enlargement of the pores, thus enhancing the

adsorption of dyes [13]. Nonetheless, too long an activation time will cause the pores to collapse and contribute to an increase in ash content [14].

Around 10,000 different dyes are produced annually from various industrial processes, and together weigh approximately 0.7 million tons [15]. In the textile industry, several varieties of dyes are used, such as reactive dyes, direct dyes, dispersed dyes, acid dyes, and basic dyes. Almost 45% of textiles produced worldwide make use of reactive dyes [16]. Remazol Brilliant Blue R (RBBR) is categorized as a reactive dye. This dye has the favourable characteristics of bright colour, a low energy consumption dyeing process and high solubility in water. The discharge of these reactive dyes in waste waters into receiving streams is highly carcinogenic and toxic to organisms [17]. Many studies have been conducted on the toxicity of dyes and their impact on the ecosystem [18]. Therefore, removal of such dyes from wastewater is very important for the environment.

Pinang (*Areca catechu*) is a tropical tree which belongs to the *Arecaceae* family. It mainly grows in a region from East Africa across tropical Asia to the central Pacific [19]. In this paper, utilization of Pinang frond to produce an activated carbon is investigated and is expected to offer a potentially cheap alternative precursor. This Pinang frond based activated carbon (PFAC) is produced via CO_2 physical activation. The effects of activation time on the PFAC's characteristics and its performance in removing RBBR dye from aqueous solution are presented.

MATERIALS AND METHODS

PFAC Preparation

Pinang frond was obtained from Sungai Petani, Kedah, Malaysia. It was cleaned and subsequently dried at 110 °C for 24 hours to remove the moisture content. The dried Pinang frond was ground and loaded into a stainless steel vertical tubular reactor placed in a tube furnace. The ramp temperature was set to 20 °C/min to achieve an activation temperature of 800°C under purified nitrogen (99.99%) at a flow rate of 150 ml/min. Once the activation temperature was reached, the activation agent of CO_2 was introduced at flow rates of 300 ml/min for 1–7 hours. The reactor was then cooled down to room temperature under nitrogen flow. The samples produced were stored in an airtight container for further characterization and adsorption studies.

Characterization of PFAC

Characteristics of the samples were analysed by using surface area analysis, scanning electron microscopy (SEM) and simultaneous thermogravimetric analysis (STA). The surface area, pore volume and average pore size of the samples were examined from the adsorption isotherms of nitrogen at 77 K by using a volumetric adsorption analyzer (ASAP 2020 Micromeritics). The BET surface area was determined using the Brunauer-Emmett-Teller (BET) equation. The total pore volumes were estimated to be the liquid volume of nitrogen at a relative pressure of 0.98. Scanning electron microscopy (SEM) analysis was used to determine the visuals of pore structure, surface structure and pore arrangement. The sample was put on carbon tape on an aluminum stub and coated with gold for electron reflection. The analysis was done by using a scanning electron microscope (Quanta 450 FEG, Netherlands). Proximate analysis was used to determine the moisture, volatile matter, fixed carbon and ash contents from the dried samples. This

analysis was carried out using a simultaneous thermal analyzer (STA) (Perkin Elmer STA 6000, USA). The STA system was interfaced to a microcomputer for data acquisition and control tasks. From the STA results, the moisture, volatile matter, fixed carbon and ash contents can be obtained, where each parameter is represented as a weight loss percentage from the total weight of the sample. The samples were heated from room temperature to 110°C in nitrogen N₂ gas until dehydration was completed to obtain the moisture content. Decomposition was applied to the sample at 900°C to determine the amount of volatile matter. Fixed carbon was obtained by switching the N₂ flow to O₂ flow. The remaining weight is represented as ash content.

Remazol Brilliant Blue R

Remazol Brilliant Blue R (RBBR) dye supplied by Sigma-Aldrich (M) Malaysia was used as an adsorbate. RBBR has a chemical formula of $C_{22}H_{16}N_2Na_2O_{11}S_3$ with a molecular weight of 626.54 g/mol. Deionized water was used to prepare all the reagents and solutions.

Batch Adsorption and Analysis System

Batch adsorption studies were carried out in a set of Erlenmeyer flasks of 250 mL with 200 mL adsorbate solution of known initial concentration. The weight of the adsorbent was fixed at 0.2 g per flask. An isothermal water bath shaker was used at fixed 120 rpm at a constant temperature. The water bath shaker was equipped with a temperature controller which can be set from 25 to 100 ± 0.1 °C and a rotation speed controller which can be fixed up to 250 ± 1 rpm. A double-beam Shimadzu UV-Visible spectrophotometer was used to measure the concentration of the adsorbates. According to Beer's law, the linear relationship between absorbance and absorbing species concentration can be written as:

$$C = \frac{A_i}{\mathscr{A}bc} \tag{1}$$

where C is the solute concentration (mg/L), A_i is the measured absorbance for component *i*, $\varepsilon \lambda$ is the molar absorptivity coefficient of solute at wavelength λ (nm), and *bc* is the path length of the cell (1 cm). The absorbance A_i was obtained by the spectrophotometer through 1 cm path length of quartz cell.

The maximum wavelength of the RBBR was 590 nm. The calibration curve for RBBR dye concentration was measured to assure the homogeneity of the absorbance reading. The calibration curve for dye was obtained from the spectrophotometer as the plot of absorbance A_i against the solute concentration C. The percentage of dye removal can be calculated by:

$$%C = \frac{(C_e - C_t)}{C_t} \times 1000$$
 (2)

where C_e and C_t in mg/L are the liquid-phase concentrations of the adsorbate at equilibrium and at any arbitrary time *t*, respectively.

Preparation of Stock and Dye Solutions

Dye powder of 1.0 g was dissolved in 1000 mL of deionized water to prepare the concentration of 1 g/L dye solution. Solutions of different initial concentrations, i.e. 50, 100, 200, 300, 400 and 500 mg/L, were prepared by a dilution process of the initial stock solution into 200 mL of deionized water.

RESULTSAND DISCUSSION

Characterization of PFAC

Characterization of PFAC is important in order to determine the properties which affect its adsorption capacity. Characterizations of activated carbon comprising the surface area characteristics, pore size distribution and surface morphology and proximate analysis need to be carried out to verify the efficiencies of an activated carbon in order to study the parameters.

Surface area and pore volume

In terms of surface area, pore volume and average pore diameter of the PFAC, the effect on it activation time were studied using the surface area analyzer. Table 1 shows that the $PFAC_{1h}$ is found to have the lowest BET surface area and total pore volume due to the lower activation time, which prevents optimum development of porosity.

Sample label	Activation time (hour)	BET surface area (m ² /g)	Total pore volume (mL/g)	Average pore diameter (nm)
Pinang frond	-	4.84	0.0087	7.20
PFAC _{1h}	1	794.75	0.4219	2.32
PFAC _{3h}	3	958.23	0.5469	2.32
PFAC _{5h}	5	873.51	0.4706	2.34
PFAC _{7h}	7	806.02	0.4331	2.35

Table 1.Surface area and pore characteristics of the PFAC.

Meanwhile, PFAC_{3h} is found to have the highest BET surface area and total pore volume, at 958.23 m²/g and 0.5469 mL/g, respectively. This result shows that this was sufficient activation time for creating a large specific surface area and high pore volume. But for PFAC_{5h} and PFAC_{7h} the surface area and total pore volume were slightly reduced compared to PFAC_{3h}, caused by the excessive activation time, which results in the wall of the pore structure becoming thin and weak, which then decreases the development of porosity. In terms of average pore diameter, all of the PFAC samples give almost the same result, i.e. 2.3 nm diameter, signifying that the PFAC is in the mesopore region.

Pore size distribution

Pore size distribution is a very important property of adsorbents which determines the fraction of the total pore volume accessible to molecules of a given size and shape. According to the classification of the International Union of Pure and Applied Chemistry (IUPAC) classification [20], the pores of absorbers can be grouped into micropores (d < 2 nm), mesopores (d = 2–50 nm) and macropores (d > 50 nm) [21]. Figure 1 illustrates the pore size distribution of the PFAC using the BJH method. From the plot, a singular sharp peak is detected in the range of 2–3 nm, which is in the mesopore region. The mesopore surface area and the mesopore volume of an activated carbon are the most important characteristics required for liquid adsorption, especially for removing dye [22].



Figure 1. Pore size distribution of PFAC using BJH method.

Surface morphology

From the SEM micrographs obtained, the surface structure and pore development on the samples can be clearly seen. Figure 2 represents the surface morphological change of the carbon material during the activation process with different pore shapes. In Figure 2(a), the surface texture of the Pinang frond precursor was rough, uneven, undulating and very few pores were produced on the surface. After the activation process, the PFAC generates more pores, as can be observed in Figure 2(b-e). Figure 2(b) for PFAC_{1h} shows that the porosities have begun to develop, but the porosity development does not look complete, as there are few pores opening and thick walls. The sample of $PFAC_{3h}$ in Figure 2(c) shows the well-organized development of a pore structure with larger opening pores, because the activation time is sufficient for porosity to develop. In samples PFAC_{5h} and PFAC_{7h} in Figure 2(d) and Figure 2(e), the pore structures were disorganized, thin and weak because the structure wall is beginning to collapse due to excess activation time. A similar phenomenon was observed in a study on the effect of activation times on coconut shell-based activated carbon, as reported by Guo et al. [14], where, with greatly increased activation time, more pores were widened, causing them to collapse. Therefore, the time should be just enough to eliminate all the moisture and cause pores to develop.



Figure 2. Scanning electron micrographs: (a) Pinang frond, (b) $PFAC_{1h}$, (c) $PFAC_{3h}$, (d) $PFAC_{5h}$ and (e) $PFAC_{7h}$.

Proximate analysis

The results for proximate analysis of the samples are tabulated in Table 2. The Pinang frond precursor was found to be high in volatile matter and moisture content. After the activation process, the volatile matter and moisture content decreased significantly, whereas the fixed carbon content increased in the sample. This condition occurs due to the influence of the pyrolytic effect at high temperature, where most of the organic substances have been degraded and discharged both as gas and liquid tars, resulting in a material with high carbon purity [23]. The higher fixed carbon content means a large carbon surface for the adsorbate to be adsorbed. However, the existence of ash may sinter and block the pores, which contributes significantly to decreasing the pores' surface area [24].

Table 2. Result from proximate analysis.

Sample	Moisture (%)	Volatile (%)	Fixed carbon (%)	Ash (%)
Pinang frond	14.42	61.30	21.20	3.08
PFAC _{1h}	6.18	20.92	75.25	2.94
PFAC _{3h}	4.54	13.28	79.74	2.44
PFAC _{5h}	4.49	13.25	79.66	2.60
PFAC _{7h}	4.40	13.19	79.62	2.79

Based on the three previous analyses for the characterization of PFAC, PFAC_{3h} gives the best result. From the surface area analyzer, PFAC_{3h} has the highest values for the BET surface area and total pore volume. In the surface morphology study, PFAC_{3h} is found to have wider opening pores as well as a more organized and stable structure. Again from the proximate analysis, high fixed carbon with low ash content is also found in the PFAC_{3h}sample. Therefore, the PFAC_{3h} sample is chosen for the batch adsorption studies.

Batch Adsorption Studies of Dye on PFAC

Figure 3 shows the RBBR removal percentage versus the time interval obtained from Eq. (2). It can be seen that the RBBR removal percentage increases with time and reaches a constant value above 20 hours. For the first 5 hours of batch adsorption, a rapid increase in RBBR removal can be observed for all RBBR concentrations. In the time interval of 5–20 hours, the RBBR removal slowly increases and reaches saturation above 20 hours. This condition is due to the fact that a large number of surface sites are available for adsorption at the initial stages, and after the passage of time, the remaining surface sites are difficult to occupy because of the repulsion between the solute molecules of the solid and bulk phases [25]. At the end (24 hours), between 40.8% and 87.6% of the RBBR has been successfully removed at all initial dye concentrations, where the latter is achieved using an initial dye concentration.



Figure 3. RBBR percentage removal versus adsorption time at various initial concentrations at 30° C on PFAC_{3h}.

CONCLUSIONS

In the present study, the preparation of activated carbon using Pinang frond from waste precursor has successfully been utilized via a physical activation method. The optimum pyrolysis and activation time of PFAC is found at 3 hours, when the BET surface area, the total pore volume and the fixed carbon content are the highest compared to other activation times. From the surface morphology, the PFAC_{3h} creates well-developed pore structures with wider pores and a stable structure. The average pore diameter and the particle size distribution of the PFAC structure are found to be in the mesopore region, which is therefore suitable to apply in a liquid phase adsorption. In the batch adsorption study, PFAC is identified as being suitable for removing RBBR dye from aqueous

solution. At the equilibrium condition, $PFAC_{3h}$ is capable of adsorbing dye and removing 87.6% of the RBBR dye at an initial concentration of 50 mg/L. Therefore, the preparation of activated carbon by applying a suitable activation time has been presented to be significant in developing the porosities and adsorption capability of the activated carbon.

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REFERENCES

- [1] Dias JM, Alvim-Ferraz M, Almeida MF, Rivera-Utrilla J, Sánchez-Polo M. Waste materials for activated carbon preparation and its use in aqueous-phase treatment: a review. Journal of Environmental Management. 2007;85:833-46.
- [2] Sumathi S, Bhatia S, Lee K, Mohamed A. Selection of best impregnated palm shell activated carbon (PSAC) for simultaneous removal of SO₂ and NOx. Journal of Hazardous Materials. 2010;176:1093-6.
- [3] Gupta V. Application of low-cost adsorbents for dye removal–A review. Journal of environmental management. 2009;90:2313-42.
- [4] Lua AC, Guo J. Activated carbon prepared from oil palm stone by one-step CO₂ activation for gaseous pollutant removal. Carbon. 2000;38:1089-97.
- [5] Herawan SG, Ahmad M, Putra A, Yusof A. Effect of Flow Rate on the Pinang Frond-Based Activated Carbon for Methylene Blue Removal. The Scientific World Journal. 2013;2013.
- [6] Bello OS, Ahmad MA, Ahmad N. Adsorptive features of banana (Musa paradisiaca) stalk-based activated carbon for malachite green dye removal. Chemistry and Ecology. 2012;28:153-67.
- [7] González P, Pliego-Cuervo Y. Physicochemical and microtextural characterization of activated carbons produced from water steam activation of three bamboo species. Journal of Analytical and Applied Pyrolysis. 2013;99:32-9.
- [8] Ahmad MA, Rahman NK. Equilibrium, kinetics and thermodynamic of Remazol Brilliant Orange 3R dye adsorption on coffee husk-based activated carbon. Chemical Engineering Journal. 2011;170:154-61.
- [9] Bouchelta C, Medjram MS, Zoubida M, Chekkat FA, Ramdane N, Bellat JP. Effects of pyrolysis conditions on the porous structure development of date pits activated carbon. Journal of Analytical and Applied Pyrolysis. 2012;94:215-22.
- [10] Herawan S, Hadi M, Ayob MR, Putra A. Characterization of activated carbons from oil-palm shell by CO₂ activation with no holding carbonization temperature. The Scientific World Journal. 2013;2013.
- [11] Nunell G, Fernández M, Bonelli P, Cukierman A. Conversion of biomass from an invasive species into activated carbons for removal of nitrate from wastewater. Biomass and Bioenergy. 2012;44:87-95.
- [12] Rashidi NA, Yusup S, Ahmad MM, Mohamed NM, Hameed BH. Activated carbon from the renewable agricultural residues using single step physical activation: a preliminary analysis. APCBEE Procedia. 2012;3:84-92.

- [13] Lua AC, Yang T. Effect of activation temperature on the textural and chemical properties of potassium hydroxide activated carbon prepared from pistachio-nut shell. Journal of Colloid and Interface Science. 2004;274:594-601.
- [14] Guo S, Peng J, Li W, Yang K, Zhang L, Zhang S, et al. Effects of CO₂ activation on porous structures of coconut shell-based activated carbons. Applied Surface Science. 2009;255:8443-9.
- [15] Senthilkumaar S, Kalaamani P, Porkodi K, Varadarajan P, Subburaam C. Adsorption of dissolved reactive red dye from aqueous phase onto activated carbon prepared from agricultural waste. Bioresource Technology. 2006;97:1618-25.
- [16] Tunc Ö, Tanacı H, Aksu Z. Potential use of cotton plant wastes for the removal of Remazol Black B reactive dye. Journal of Hazardous Materials. 2009;163:187-98.
- [17] Baskaralingam P, Pulikesi M, Ramamurthi V, Sivanesan S. Modified hectorites and adsorption studies of a reactive dye. Applied clay science. 2007;37:207-14.
- [18] Vadivelan V, Kumar KV. Equilibrium, kinetics, mechanism, and process design for the sorption of methylene blue onto rice husk. Journal of Colloid and Interface Science. 2005;286:90-100.
- [19] Staples GW, Bevacqua RF. Areca catechu (betel nut palm): Species Profiles for Pacific Island Agroforestry. 2006.
- [20] Sing KS. Reporting physisorption data for gas/solid systems with special reference to the determination of surface area and porosity (Recommendations 1984). Pure and Applied Chemistry. 1985;57:603-19.
- [21] Deng H, Yang L, Tao G, Dai J. Preparation and characterization of activated carbon from cotton stalk by microwave assisted chemical activation— application in methylene blue adsorption from aqueous solution. Journal of Hazardous Materials. 2009;166:1514-21.
- [22] Wei ITA. Preparation, characterization and evaluation of mesoporous activated carbons derived from agricultural by-products for adsorption of methylene blue and 2,4,6- trichlorophenol. Malaysia: Universiti Sains Malaysia; 2008.
- [23] Ahmad M, Wan Daud W, Aroua M. CO2/CH4 and O2/N2 kinetic selectivities of oil palm shell-based carbon molecular sieves. Journal of Oil Palm Research. 2008;20:453-60.
- [24] Huang Y, JIN B, Zhong Z, Zhong W, Xiao R. Characteristic and mercury adsorption of activated carbon produced by CO₂ of chicken waste. Journal of Environmental Sciences. 2008;20:291-6.
- [25] Khattri S, Singh M. Removal of malachite green from dye wastewater using neem sawdust by adsorption. Journal of Hazardous Materials. 2009;167:1089-94.