

ORIGINAL ARTICLE

A review of wastewater bacterial bio oxidation: mechanisms, reactions, and behaviors

S. Al-Amshawee^{1,*} and M. Y. M. Yunus^{1,2}

¹Faculty of Chemical and Process Engineering Technology, Universiti Malaysia Pahang, Gambang 26300, Malaysia. ²Earth Resource and Sustainability Centre (ERAS), Universiti Malaysia Pahang, Gambang 26300, Malaysia.

ABSTRACT – The most overlooked biological process is the oxidation process, due to the poor technical understanding of bioremediation and the many unexplained biological mechanisms. As a matter of fact, wastewater companies favor the physical and chemical approaches over the biological means to ensure process performance. Biological oxidation provides environmentally friendly treatment with limited operational requirements. It has a tremendous ability to metabolize ammonia, H₂S, phosphate, and ferrous iron under the right conditions. Bacterial communities attack these wastewater components through using oxygen as a strong electron acceptor, resulting in simplified forms of wastewater components (e.g., ammonia to nitrogen gas). This manuscript reviews in details the mechanisms of wastewater bacteria against various pollutants, with the possible reactions and behaviors. It also presents the benefits of utilizing wastewater microorganisms against different pollutants.

ARTICLE HISTORY

Received: 10 Aug. 2020 Revised: 24 Sep. 2020 Accepted: 30 Sep. 2020

KEYWORDS

Biological oxidation Wastewater treatments Nitrification Denitrification H₂S

INTRODUCTION

Discharge and dumping of chemically contaminated wastewater, including dyes, pesticides, solvents, and heavy metals threats our freshwater [1]. Scientific understanding of the wastewater impact has evolved with good credibility to prove several a number of severe damages triggered by contaminated water. Various quantities of contaminants have been found in very remote locations from their discharge station [2]. As a result, a scientific approach has begun to understand and assess the ability of different industries to produce profits without harm [3–4].

It is important to define the relationship between the sort of bacteria and the obtainable performance because the bacterial metabolism controls the biotreatment efficacy. Most biotreatments are derived from engineering understanding, and the bacterial dynamics are largely neglected. A major approach in wastewater treatments (WWTs) will be developed once the microbial dynamics are considered. Furthermore, environmental conditions are the main factors that have revealed the changes in bacterial communities [5–7].

Wastewater bacteria

The amount of heterotrophic and autotrophic bacteria varies in each wastewater on the basis of the treatment system and the sort of wastewater used [8]. Betaproteobacteria is considered to be the most densely populated organism in municipal wastewater, while Phylum Proteobacteria is the predominant microorganism, and *Bacteroidetes*, *Acidobacteria* and *Chloroflexi* are subdominant [9–10]. *Tetrasphaera*, *Trichococcus*, *Candidatus Microthrix*, *Rhodoferax*, *Rhodobacter*, *Hyphomicrobium*, p-55-a5, P2CN44 and B45 bacteria are the most frequently identified, according to the evaluation of 20 wastewater treatment plants [11]. Approximately 6.3%-7.4% of wastewater microorganisms are *Ascomycota*, while *Euryarcheota* is recognized as predominant with 1.5% of total wastewater microorganisms [12]. The wastewater remediation always favors some microorganism classes over the rest, according to the molecular studies. The availability of molecular data may support the scientific understanding of wastewater bioremediation (e.g., anaerobic ammonium oxidation (anammox), nitrification). Oxidation ditches, and membrane bioreactors (MBR) have been found to have fewer microscopic species than anaerobic/anoxic/oxic (A2O) and anaerobic/oxic (AO) processes [9,13]. In addition, low diversity of microorganisms in MBR could be due to the low availability of biodegradable organics and long solid retention time (SRT) [9].

Much of the available data on wastewater treatment is focused on scientific study [14]. The aerobic granulation method is a strong example among biological treatments due to the high population of microorganisms such as *Thauera* sp., *Aquabacterium* sp. and *Flavobacterium* sp. These microorganisms support their attachment by creating extracellular polymer substance (EPS), and proceed the denitrification process with slow evolution [15–17]. Certain microbial groups such as filamentous can be encouraged to expand under ample capital of energy-rich substrates such as fructose and glucose [18]. Domestic WWTs have been found to have a higher diversity of ammonia oxidizing bacteria (AOB) than municipal WWTs, and the major bacteria in the nitrification cycle are Nitrosomonas sp. [19–20]. Another great example of bioremediation is the sequencing bacter (SBR) that generates high biodiversity and population through its pretty

decent engineering. Bioremediation failure in WWTs is due to poor engineering understanding of biological treatment through providing unsatisfactory circumstances such as high range of chemical inhibitors, low DO, extreme pH, and low temperatures [21].

According to a survey conducted on 14 WWTs, approximately 95% of the activated sludge composition comprises of *Myxococcales, Planctomycetales, Clostridiales, Verrucomicrobiales, Xanthomonadales, Rhizobiales, Burkholderiales, Rhodocyclales, Anaerolineales,* and *Sphingobacteriales* [10]. Microscopic species differ based on the geographical place, and circumstances like temperature. For instance, *Chloroflexi, Actinobacteria,* and *Betaproteobacteria* comprise 50% of the total microbial population regardless of temperature, while *Alphaproteobacteria,* and *Saprospiraceae* as a whole are affected by temperature levels [22]. In the same way, filamentous bacteria have been detected varying by type of WWT, and their geographical location. Usually, wastewater contains a small amount of filamentous bacteria that enable the formation of a microbial matrix. Despite this, an excessive number of filamentous bacteria leads to sludge bulking. Wastewater often involves different filamentous groups, such as *Microthrix parvicella*, Type 1863 *Acinetobacter, Mycobacterium fortuitum*, and *Nostocola limicola* I and II, which make up 1.86%-8.99% of the total population of wastewater microorganisms [23].

The usual activity in wastewater is the development of an immobilized multilayer matrix of microorganisms on solid surfaces utilizing hydrophobic interactions, covalent bonds, and electrostatic interactions known as biofilm formation [16]. These microbial communities include EPS, cell wall components, *cilia*, and *fimbriae*. In addition, the fine engineering design of the biofilm structure provides oxygen penetration through determined depth, mass diffusion, and genetic transfer through multiple channels and pores, leading to toxic compound resistance, metabolic activity, and bacterial growth [24–25]. EPS compositions are based on cell lysis, material shedding from the cell surface, and material absorption from the surrounding area. A researcher stated that the structure of EPS differs between biofilms and activated sludge in the same environment [26]. Free nitrous acid and ammonia in WWTs are the key causes for the microbial complex formation of biofilm bacteria to protect against harmful effects [27]. EPS structure provides a diffusion mechanism that reduces the nearby toxic level. The constitute of EPS is about 80-95% organic biomass, while the rest is bacteria [28].

Bacterial Bio Oxidation

Bio oxidation is the most environmentally sustainable approach for the removal of phosphorus (P), nitrogen (N) and organic matters from wastewater. In order to avoid aquatic eutrophication, the bio oxidation mechanism is important for wastewater. In fact, many WWTs rely on microbial activities for the removal of pollutants. Aerobic systems have been found to be unable to minimize P and N, but have high rates of organic carbon removal. A conventional WWT system was used to deliver different amounts of dissolved oxygen to trigger all bacterial groups for simultaneous nitrification, denitrification and phosphate removal (SNDPR) [29–30]. Ammonium oxidizing bacteria (e.g., *Nitrosomonas*) and nitrite oxidizing bacteria (e.g., *Nitrospira*) [complete ammonia oxidizing bacteria (comammox)] convert ammonia to nitrite through aerobic conditions through a nitrification process [31]. Anoxic conditions are then used to convert nitrite to N_2 by the denitrification process. At the same time, there are different microbial groups to treat polyP known as polyphosphate (polyP) accumulating organisms (PAOs) [32]. During anaerobic conditions, PAO uses polyP at the intracellular as food, accumulates phosphate from wastewater, and seizes polyP in cells under aerobic conditions.

A researcher examined bioremediation and found that inorganic chemicals were used as energy source for aerobic bacteria (nitrification) under aerobic circumstances, followed by heterotrophic and anaerobic bacteria producing biomass through anaerobic or anoxic conditions (denitrification) [33]. Xue et al. [34] confirmed that dissolved oxygen (DO) determines the overall efficacy of biodegradation and relates to first-order biodegradation constants. Some elements and compounds, on the other hand, resist biodegradation at any DO concentration [33]. Shortly, AOB (ammonia oxidizing bacteria), and NOB (nitrite oxidizing bacteria) are heterotrophs, while PNA (partial nitritation annamox) is thought to be heterotrophic. Figure 1 shows the full ammonia conversion cycle by PNA [35].

Although bio oxidation is considered to be an eco-friendly process between other treatments, it emits CH₄ through anaerobic digestion, CO₂ through respiration, and N₂O through nitrification and the denitrification [36]. A small volume of N₂O emissions is equivalent to a large carbon footprint, and its impact is 265 times greater than CO₂, where the lifetime of N₂O gas is 114 years in the atmosphere [37]. In fact, 26% of global warming is caused by N₂O pollution [38]. Heterotrophic denitrification, denitrification of nitrifiers, and oxidation of hydroxylamine (NH₂OH) are the main sources of N₂O emissions (see Figure 2) [39]. Figure 2 demonstrates the process of production of N₂O by AOB (i.e., nitrifiers' denitrification, and NH₂OH oxidation), and heterotrophic denitrifiers (i.e., heterotrophic denitrification) [39].

A wastewater treatment with low temperatures, high concentration of NO_2^- , and low concentration of DO are the key factors behind the N₂O release from the biological cycle. At these conditions, the AOB variety overcomes the NOB population, especially during low DO ranges. Chen et al. [40] found that the emission of N₂O was due to the oxic phase of AOB. Converting the anaerobic phase to idle conditions will reduce 42% of N₂O emissions due to the consumption of N₂O by heterotrophic denitrification. Ahn et al. [41]confirmed, through their empirical study, that NO and N₂O were high ranges at low DO levels. When DO levels rise from 1 to 4.5 mg/l, N₂O decreases from 6% to 2.2%, according to Pijuan et al. [42].



Figure 1. Illustration of PNA reaction including ammonia [35].



Figure 2. Illustration of processes for N₂O generation [38].

Ammonia bio oxidation

High levels of ammonia concentrations can turn water into a toxic medium for aquatic life, because it makes a high demand for oxygen, resulting in eutrophication. Ammonia oxidizing bacteria (AOB) and ammonia oxidizing archaea (AOA) population are determined by the influent concentration of ammonia. AOA conducts an autotrophic oxidation cycle in WWTs. For example, the AOB population in industrial WWTs is higher than AOA due to high ammonia availability, whereas the AOA abundance is higher than AOB in municipal WWTs [43]. However, a well-removal process can be achieved even under a low population of autotrophic nitrifiers.

Wang et al. [44] and Ma et al. [45] observed that heterotrophic nitrifiers have been developed from the *Azoarcus*, *Paracoccus*, *Thauera*, and *Comamonas* genera for the remediation of rich ammonium coking wastewater by activated sludge reactors. Nitrifiers and denitrifiers often request a variety of substrates for their production. In the process of rotating biological contactors (RBC), it was found that the population of nitrifers increased at low abundance of denitrifers (*Azoarcus, Thauera, Paracoccus*, and *Rhodanobacter* genera) [46]. It was observed that the AOB resided on the flocs and the outer surfaces of granules, while the anammox bacteria remained in the inner zone [47]. AOB partially oxidizes ammonia to NO_2^- , while the remaining NH_4^+ is oxidized by anammox using NO_2^- as an electron acceptor [38].

The DNA testing for denitrifiers showed that most of the AOBs belong to *Nitrosospira* sp., and *Nitrosomonas* sp., and share various similarities [12]. Denitrifier functions are influenced by the same circumstances as DO, N, organic load, and temperature with varying outputs. It has long been belived that oxic conditions are suitable mediums for ammonia oxidation, but Broda has suspected the presence of chemolithoautotrophic anammox bacteria for two decades on the basis of thermodynamic calculations [48]. The electron acceptor was nitrite in the ammonia oxidation process [36], [49].

AOB delivers nitritation by converting ammonium to nitrite, then the NOB group delivers nitratation by converting nitrite to nitrate. Nitrification concept applies to both nitritation and nitratation, whereas AOB, and NOB groups are called nitrifiers [38]. Often the process of nitrification is known as the method of using NO_2^- to convert NH_4^+ to NO_3^- by microorganisms of AOB and NOB. Hydroxylamine (NH₂OH) is usually produced as an intermediate product when NH_4^+ is oxidized with one of O_2 atoms [38]. The hydroxylamine oxidoreductase then oxidizes the intermediate to NO_2^- [50]. The process of decomposition of the intermediate produces N_2O . Sometimes, N_2O can be produced by the denitrification process at excessive concentrations of nitrite, where N_2O , and N_2 are formed by the reduction of NO_2 [51-52].

Oxygen is required for the bioremediation process to be used as an electron acceptor, and as a substrate for the production of hydroxylamine by NH_4 oxidation. 4.2 g of oxygen is required for each gram of ammonium nitrogen nitrified by the nitrification process [53]. Nitrite may be used as an electron acceptor in the case of low DO concentration.

A broken reaction causes serious ecological risks, such as foam formation, methemoglobinemia, toxic nitrate and/or nitrite, and the death of fish [54]. Nitrification and denitrification were studied in a number of WWTs, such as aeration tanks, moving bed biofilm reactor (MBBR), and rotating disk reactor [55–57]. Figure 3 displays the oxidation process.



Figure 3. Overview of the oxidation process [58].

The TKN concentration is a crucial parameter to present the achievement of nitrification. Reactions that have occurred through the nitrification and denitrification processes are outlined in equations 1, 2, 3, and 4. These conventional reactions have been identified for a long time [49].

$NH_4^+ + 1.5O_2 \rightarrow NO_2^- + 2H^+ + 2H_2O$	(1)
$NO_2^- + 0.5O_2 \rightarrow NO_3^-$	(2)
$2NO_3^- + 10H^+ + 10e^- \rightarrow N_2 + 2OH^- + 4H_2O$	(3)
$2NO_2^- + 6H^+ + 6e^- \rightarrow N_2 + 2OH^- + 2H_2O$	(4)
	• .1 1

As far as denitrification is concerned, 2.47 g of methanol is needed for each nitrate nitrogen gram, in case methanol is used as an electron donor [54]. The right electron donor is methane because it is readily accessible and cheap for denitrification, but it is slowly consumed [59]. A large amount of organic carbon is also required to provide a complete process. Ammonium oxidation can be occurred under anoxic conditions using organic compounds and hydrogen as an electron donors, according to Schmidt and Bock (1997) [60], [61]. Nitrite is the perfect electron acceptor for the oxidation process (see equations 5 and 6).

$5NH_4^+ + 3NO_3^- \rightarrow 4N_2 + 9H_2O + 2H^+$	(5)
$NH_4^+ + NO_2^- \rightarrow N_2 + 2H_2O$	(6)

Approximately 10% of the feed ammonia and nitrite have been converted to NO_3^- under anaerobic conditions, and the main product is still N_2 . Equation 7 describes the total balance of nitrogen from the feed to the last product via the nitrification [62].

$$NH_{4}^{+} + 1.31NO_{2}^{-} + 0.0425CO_{2} \rightarrow 1.045N_{2} + 0.22NO_{3}^{-} + 1.87H_{2}O + 0.09OH^{-} + 0.0425CH_{2}O \text{ (Biomass)}$$
(7)
The overall balance of anammox cultures was estimated as shown in equation 8.

 $NH_4^+ + 1.31NO_2^- + 0.066HCO_3^- + 0.13H^+ \rightarrow 1.02N_2 + 0.26NO_3^- + 0.066CH_2O_{0.5}N_{0.15} + 2.03H_2O$ (8)

Phosphorus removal

Polyphosphate accumulating organisms (PAOs) are used in WWTs to remove phosphate through aerobic or anaerobic conditions. PAOs are capable of eliminating phosphate by using nitrate or nitrite as electron acceptors [63]. *Accumulibacter* sp., and *Tetrasphaera* sp. are examples on the PAOs. *Accumulibacter* sp. consume volatile fatty acids and store polyhydroxyacids (PHAs) while *Tetrasphaera* sp. take amino acids and do not store PHAs. 3-10% of the population is *Accumulibacter* sp., while 30-35% is *Tetrasphaera* sp. [64]. The carbon to phosphate ratio determines the abundance of *Accumulibacter* sp., and *Tetrasphaera* sp. [65]. *Dechloromonas* is another group of PAOs which is capable of nitrite/nitrate reduction, PHA storage, polyphosphate, and acetate acceptance [66]. *Candidatus Accumulibacter phosphatis* is the most known group of PAOs belonging to *Rhodocyclaceae* family of the Betaproteobacteria class. Figure 4 shows an examination result from full scale oxidation WWT in Japan of finding the working microorganisms on P removal.



Figure 4. Distribution of Phylogenetic's gene clones in WWT oxidation ditch [67].

H₂S oxidation

The common species for sulfide removal are purple sulfur bacteria, colorless sulfur bacteria, and phototrophic sulfur bacteria. As wastewater has a high level of nitrates under denitrifying conditions, the sulfur bacteria use nitrates as an electron acceptor. The oxidation process of H₂S produces sulphate and sulfur. Sulfide is then oxidized by reducing nitrate into nitrite by the sulfide oxidizing bacteria (SOB). The sulfide oxidation reaction is zero-order. In the event of incomplete oxidation of sulfide, the solution becomes white due to the excess amount of the intermediate element of the reaction known as sulfur. The suggested incomplete reaction is shown in equation 9. NO₃⁻ + HS⁻ \rightarrow S⁰ + NO₂⁻ + OH⁻ (9)

Sublette and Sylvester (1987) reported a practical oxidation reaction by converting 40%-60% of the sulfide fed to sulphate [68]. Possible complete reactions are presented in equation 10 and 11.

$\mathrm{HS}^{-} + 1/2\mathrm{O}_2 \rightarrow \mathrm{S}^0 + \mathrm{OH}^{-}$	(10)
$NO_3^- + H^+ + 2e^- \rightarrow NO_2^- + OH^-$	(11)

Iron oxidation

Ferrous iron acts as an electron donor under anaerobic conditions, or as an additional electron donor in the presence of acetate. In addition, iron is the most suitable metal for the redox process [16]. Iron-reducing bacteria also treat ferric iron in sediments and soils under anoxic conditions [69]. Light can be used as an energy source with O_2 for phototrophic bacteria to carry out biological oxidation by taking the electron from the iron as an electron donor and giving it to nitrate as an electron acceptor [70]. Equation 12 shows the possible oxidation reaction.

 $10 \text{ FeCO}_3 + 2 \text{ NO}_3^- + 24 \text{ H}_2\text{O} \rightarrow 10 \text{ Fe}(\text{OH})_3 + \text{N}_2 + 10 \text{ HCO}_3^- + 8\text{H}^+$ (12)

Successful experiments on the use of ferrous iron to reduce nitrous oxide and nitrite have been reported [71-72]. Finally, the work in the present review has led the authors to illustrate the complete bio oxidation of ammonia, phosphate and polyphosphate, and H_2S (see Figure 5).



Figure 5. Flowchart of bio oxidation. (a) Ammonia bio oxidation; (b) Phosphate and polyphosphate bio oxidation; (c) H₂S bio oxidation.

Antibiotics-resistant bacteria

Ineffective degradation leads to the growth and development of antibiotic-resistant bacteria (ARB) in the environment [49]. The main home for the evolution of ARB is wastewater, which is a perfect culture for the production of antibiotic resistance genes (ARGs). As a result, the mineralization processes of wastewater pollutants by microbes are quite important to reveal the evolution of ARGs.

During various wastewater treatments, the ARB population and diversity were found to be the highest in the wastewater influent, followed by treatment effluent, anaerobic digestion sludge, and activated sludge, depending on the concentration of wastewater pollutants. Sludge treatment is usually unable to reduce the population and growth of ARBs, while other wastewater treatments are likely to result in more than 99% removal [73]. For this end, a disinfection process is needed to ensure the removal of ARBs. However, some methods may cause reactivation and regeneration of ARBs, such as chlorination [74]. In addition, the concentration and type of antibiotics within the influent determines the effluent bacterial diversity. For instance, bacteria from *Clostridia* and *Bacilli* classes, *Firmicutes*, and phyla Proteobacteria predominated effluents after treating penicillin-containing wastewater [75].

It has also been reported that the population of *Epsilonproteobacteria* grows proportionally with the presence of triclosan, quinolones, sulfonamides, penicillin, and tetracyclines, while these components have a negative affect on the growth of *Firmicutes*, and *Gammaproteobacteria* [76]. Another study showed a high diversity and abundance of different genes during the treatment of tetracycline wastewater, such as *Rhodobacter*, *Novosphingobium*, *Paracoccus*, *Longilinea*, *Azonexus*, *Hyphomicrobium*, *Prosthecobacter*, *Armatimonas*, and *Sulfuritalea* which led to a reduction in non-tetracycline ARGs [77].

Generally, dye-containing wastewater reflects sunlight from penetrating inside the reactor. Thus, multiple types of microbes will be idled, due to the depletion of sunlight. As a result, the discharge of polluted wastewater threatens our healthy life, and the ecosystem by increasing severe risks, and serious damages.

CONCLUSION

The disposal of wastewater continues to contaminate waterways, notwithstanding the rules and legislations for the discharge of effluents. Scientists have been developing the biological oxidation data, but it is slowly occurring. For the time being, adequate and reliable knowledge on bioremediation is available to structure a technique that utilises wastewater microbes in maximum effort to bio decompose suspended contaminants. The oxidation process map incorporates much of the existing evidence and demonstrates a significant contribution to the advancement of current understanding. One of the interesting points in this research is that bacterial communities and families do teamwork during bioremediation. The operational circumstances must therefore satisfy the dominant or most of the bacteria used.

REFERENCES

- [1] S. Al-amshawee and Y. Yunus, "Influence of Light Emitting Diode (LED) on Microalgae," J. Chem. Eng. Ind. Biotechnol., vol. 5, no. 2, pp. 9–16, 2019, doi: 10.15282/jceib.v5i2.3771.
- [2] I. Oller, S. Malato, and J. A. Sánchez-Pérez, "Combination of Advanced Oxidation Processes and biological treatments for wastewater decontamination-A review," *Sci. Total Environ.*, vol. 409, no. 20, pp. 4141–4166, Sep. 2011, doi: 10.1021/es5017087.
- [3] A. Hildebrandt, S. Lacorte, and D. Barceló, "Sampling of water, soil and sediment to trace organic pollutants at a river-basin scale," pp. 1075–1088, 2006, doi: 10.1007/s00216-006-0486-2.
- [4] D. Barceló and M. Petrovic, "Emerging contaminants in wastewaters," *TrAC Trends in Analytical Chemistry*, vol. 26, no. 11. p. 1019, 2007, doi: 10.1016/j.trac.2007.10.006.
- [5] F. Ju and T. Zhang, "Bacterial assembly and temporal dynamics in activated sludge of a full-scale municipal wastewater treatment plant," *ISME J.*, vol. 9, no. 3, pp. 683–695, 2015, doi: 10.1038/ismej.2014.162.
- [6] S. Al-amshawee, M. Yusri, B. Mohd, R. Bin, M. Yunus, and J. G. Lynam, "Journal of Water Process Engineering Zero waste system comprised of fixed bed biofilm reactor, ozone oxidation, and electrodialysis desalination for wastewater sustainability," J. Water Process Eng., vol. 38, pp. 101593, 2020, doi: 10.1016/j.biortech.2016.08.031.
- [7] A. A. Sajjad, Y. H. Teow, and A. W. M. Hussain, "Sustainable approach of recycling palm oil mill effluent (POME) using integrated biofilm/membrane filtration system for internal plant usage," J. Teknol., vol. 80, no. 4, pp. 165–172, 2018, doi: 10.11113/jt.v80.11054.
- [8] A. Cydzik-Kwiatkowska and M. Zielińska, "Microbial composition of biofilm treating wastewater rich in bisphenol A," J. Environ. Sci. Heal. Part A Toxic/Hazardous Subst. Environ. Eng., vol. 53, no. 4, pp. 385–392, 2018, doi: 10.1080/10934529.2017.1404326.
- [9] M. Hu, X. Wang, X. Wen, and Y. Xia, "Bioresource Technology Microbial community structures in different wastewater treatment plants as revealed by 454-pyrosequencing analysis," *Bioresour. Technol.*, vol. 117, pp. 72–79, 2012, doi: 10.1016/j.biortech.2012.04.061.
- [10] X. Wang, M. Hu, Y. Xia, X. Wen, and K. Ding, "Pyrosequencing Analysis of Bacterial Diversity in 14 Wastewater Treatment Systems in China," 2012, doi: 10.1128/AEM.01617-12.
- [11] S. J. McIlroy *et al.*, "MiDAS: The field guide to the microbes of activated sludge," *Database*, vol. 2015, 2015, doi.org/10.1093/database/bav062.
- [12] X. Wang, Y. Xia, X. Wen, Y. Yang, and J. Zhou, "Microbial Community Functional Structures in Wastewater Treatment Plants as Characterized by GeoChip," vol. 9, no. 3, pp. 1–10, 2014, doi: 10.1371/journal.pone.0093422.
- [13] S. Al-Amshawee *et al.*, "Electrodialysis Desalination for Water and Wastewater: A Review," *Chem. Eng. J.*, vol. 380, p. 122231, 2019, doi: 10.1016/j.cej.2019.122231.
- [14] M. Pronk, M. K. De Kreuk, B. De Bruin, P. Kamminga, R. Kleerebezem, and M. C. M. Van Loosdrecht, "Full scale performance of the aerobic granular sludge process for sewage treatment," *Water Res.*, 2015, doi: 10.1016/j.watres.2015.07.011.
- [15] J. Li, L. Bin Ding, A. Cai, G. X. Huang, and H. Horn, "Aerobic sludge granulation in a full-scale sequencing batch reactor," *Biomed Res. Int.*, vol. 2014, 2014, doi: 10.1155/2014/268789.
- [16] S. Al *et al.*, "Biocarriers for biofilm immobilization in wastewater treatments : a review," *Environ. Chem. Lett.*, no. 2006, 2020, doi: 10.1007/s10311-020-01049-y.
- [17] S. K. Al-Amshawee, M. Y. Yunus, and A. A. Azoddein, "A Novel Microbial Biofilm Carrier for Wastewater Remediation," *IOP Conf. Ser. Mater. Sci. Eng.*, vol. 736, no. 7, 2020, doi: 10.1088/1757-899X/736/7/072006.
- [18] B. Y. Moy, J. Tay, S. Toh, Y. Liu, and S. T. Tay, "High organic loading influences the physical characteristics of aerobic sludge granules," pp. 407–412, 2002, doi: 10.1046/j.1472-765X.2002.01108.x.
- [19] T. Zhang, L. Ye, A. H. Y. Tong, M. F. Shao, and S. Lok, "Ammonia-oxidizing archaea and ammonia-oxidizing bacteria in six full-scale wastewater treatment bioreactors," *Appl. Microbiol. Biotechnol.*, vol. 91, no. 4, pp. 1215–1225, 2011, doi: 10.1007/s00253-011-3408-y.
- [20] S. K. Al-Amshawee, M. Y. Yunus, and A. A. Azoddein, "A Review on Hybrid Processes for Palm Oil Mill Effluent: Possible Approaches," *IOP Confer. Ser. Mater. Sci. Eng.*, vol. 736, no. 2, 2020, doi.org/10.1088/1757-899X/736/2/022036.
- [21] J. I. Prosser, Autotrophic Nitrification in Bacteria, Adv. Microb. Physiol., vol. 30, no. (C), pp. 125–181. 1989, doi.org/10.1016/S0065-2911(08)60112-5.
- [22] A. Muszyński, A. Tabernacka, and A. Miłobedzka, "Long-term dynamics of the microbial community in a full-scale wastewater treatment plant," *Int. Biodeterior. Biodegrad.*, vol. 100, no. 100, pp. 44–51, May 2015, doi: 10.1016/j.ibiod.2015.02.008.
- [23] F. Guo and T. Zhang, "Profiling bulking and foaming bacteria in activated sludge by high throughput sequencing," *Water Res.*, vol. 46, no. 8, pp. 2772–2782, 2012, doi: 10.1016/j.watres.2012.02.039.
- [24] Y. Cohen, "Biofiltration The treatment of fluids by microorganisms immobilized into the filter bedding material: a review," *Bioresour. Technol.*, vol. 77, no. 3, pp. 257–274, 2001, doi: 10.1016/S0960-8524(00)00074-2.
- [25] M. Shuler, F. Kargi, and F. Kargi, "Bioprocess engineering: basic concepts," 2002, www.sidalc.net/cgibin/wxis.exe/?IsisScript=LIBRO.xis&method=post&formato=2&cantidad=1&expresion=mfn=02899.

- [26] B. Mahendran, L. Lishman, and S. N. Liss, "Structural, physicochemical and microbial properties of flocs and biofilms in integrated fixed-film activated sludge (IFFAS) systems," *Water Res.*, vol. 46, no. 16, pp. 5085–5101, 2012, doi: 10.1016/j.watres.2012.05.058.
- [27] A. Cydzik-Kwiatkowska, K. Bernat, M. Zielińska, and I. Wojnowska-Baryła, "Cycle length and COD/N ratio determine properties of aerobic granules treating high-nitrogen wastewater," *Bioprocess Biosyst. Eng.*, vol. 37, no. 7, pp. 1305–1313, 2014, doi: 10.1007/s00449-013-1102-4.
- [28] A. Raszka, M. Chorvatova, and J. Wanner, "Review The role and significance of extracellular polymers in activated sludge . Part I: Literature review," pp. 411–424, 2006, doi: 10.1002/aheh.200500640.
- [29] C. Li, S. Liu, T. Ma, M. Zheng, and J. Ni, "Simultaneous nitrification, denitrification and phosphorus removal in a sequencing batch reactor (SBR) under low temperature," *Chemosphere*, vol. 229, pp. 132–141, 2019, doi: 10.1016/j.chemosphere.2019.04.185.
- [30] S. K. Al-Amshawee, M. Y. Yunus, and A. A. Azoddein, "A Review on Aerobic Biological Processes for Palm Oil Mill Effluent: Possible Approaches," *IOP Confer. Ser. Mater. Sci. Eng.*, vol. 736, no. 2, 2020, doi: 10.1088/1757-899X/736/2/022035.
- [31] M. A. H. J. Van Kessel *et al.*, "Complete nitrification by a single microorganism," *Nature*, vol. 528, no. 7583, pp. 555–559, 2015, doi: 10.1038/nature16459.
- [32] Z. Yuan, S. Pratt, and D. J. Batstone, "Phosphorus recovery from wastewater through microbial processes," *Curr. Opin. Biotechnol.*, vol. 23, no. 6, pp. 878–883, 2012, doi: 10.1016/j.copbio.2012.08.001.
- [33] S. Suarez, J. M. Lema, and F. Omil, "Removal of Pharmaceutical and Personal Care Products (PPCPs) under nitrifying and denitrifying conditions," *Water Res.*, vol. 44, no. 10, pp. 3214–3224, 2010, doi: 10.1016/j.watres.2010.02.040.
- [34] W. Xue, C. Wu, K. Xiao, X. Huang, and H. Zhou, "Elimination and fate of selected micro-organic pollutants in a full-scale anaerobic/anoxic/aerobic process combined with membrane bioreactor for municipal wastewater reclamation," *Water Res.*, vol. 44, no. 20, pp. 5999–6010, 2010, doi: 10.1016/j.watres.2010.07.052.
- [35] D. R. Speth, M. H. In'T Zandt, S. Guerrero-Cruz, B. E. Dutilh, and M. S. M. Jetten, "Genome-based microbial ecology of anammox granules in a full-scale wastewater treatment system," *Nat. Commun.*, vol. 7, p. 11172, 2016, doi: 10.1038/ncomms11172.
- [36] G. Mannina *et al.*, "Science of the Total Environment Greenhouse gases from wastewater treatment A review of modelling tools," *Sci. Total Environ.*, vol. 551–552, pp. 254–270, 2016, doi: 10.1016/j.scitotenv.2016.01.163.
- [37] IPCC, Climate Change 2007 The Physical Science Basis: Working Group I Contribution to the Fourth Assessment Report of the IPCC (Climate Change 2007), 2007.
- [38] T. Maria, S. Malamis, A. Guisasola, J. Antonio, C. Noutsopoulos, and E. Katsou, "Science of the Total Environment A review on nitrous oxide (N 2 O) emissions during biological nutrient removal from municipal wastewater and sludge reject water," *Sci. Total Environ.*, vol. 596–597, pp. 106–123, 2017, doi: 10.1016/j.scitotenv.2017.03.191.
- [39] B. J. Ni and Z. Yuan, "Recent advances in mathematical modeling of nitrous oxides emissions from wastewater treatment processes," *Water Research*, vol. 87. pp. 336–346, 2015, doi.org/10.1016/j.watres.2015.09.049.
- [40] Y. Chen, D. Wang, X. Zheng, X. Li, L. Feng, and H. Chen, "Chemosphere Biological nutrient removal with low nitrous oxide generation by cancelling the anaerobic phase and extending the idle phase in a sequencing batch reactor," *Chemosphere*, vol. 109, pp. 56–63, 2014, doi: 10.1016/j.chemosphere.2014.02.011.
- [41] J. H. Ahn and K. Chandran, "Comparison of Partial and Full Nitrification Processes Applied for Treating High-Strength Nitrogen Wastewaters: Microbial Ecology through Nitrous Oxide Production," pp. 2734–2740, 2011, doi: 10.1021/es103534g.
- [42] M. Pijuan, J. Torà, A. Rodríguez-Caballero, E. César, J. Carrera, and J. Pérez, "Effect of process parameters and operational mode on nitrous oxide emissions from a nitritation reactor treating reject wastewater," *Water Res.*, vol. 49, no. 1, pp. 23–33, 2014, doi: 10.1016/j.watres.2013.11.009.
- [43] Y. Bai, Q. Sun, D. Wen, and X. Tang, "Abundance of ammonia-oxidizing bacteria and archaea in industrial and domestic wastewater treatment systems," pp. 1–8, 2012, doi: 10.1111/j.1574-6941.2012.01296.x.
- [44] Z. Wang *et al.*, "Abundance and Diversity of Bacterial Nitrifiers and Denitrifiers and Their Functional Genes in Tannery Wastewater Treatment Plants Revealed by High- Throughput Sequencing," pp. 1–19, 2014, doi: 10.1371/journal.pone.0113603.
- [45] Q. Ma *et al.*, "Bacterial community compositions of coking wastewater treatment plants in steel industry revealed by Illumina high-throughput sequencing," *Bioresour. Technol.*, vol. 179, pp. 436–443, 2015, doi: 10.1016/j.biortech.2014.12.041.
- [46] X. Peng, F. Guo, F. Ju, and T. Zhang, "Shifts in the microbial community, nitrifiers and denitrifiers in the biofilm in a fullscale rotating biological contactor," *Environ. Sci. Technol.*, vol. 48, no. 14, pp. 8044–8052, 2014, doi: 10.1021/es5017087.
- [47] Z. rui Chu, K. Wang, X. kun Li, M. ting Zhu, L. Yang, and J. Zhang, "Microbial characterization of aggregates within a onestage nitritation-anammox system using high-throughput amplicon sequencing," *Chem. Eng. J.*, vol. 262, pp. 41–48, 2015, doi: 10.1016/j.cej.2014.09.067.
- [48] E. Broda, "Two kinds of lithotrophs missing in nature," Z. Allg. Mikrobiol., vol. 17, no. 6, pp. 491–493, 1977, doi: 10.1002/jobm.19770170611.
- [49] A. Cydzik-kwiatkowska and M. Zielin, "Bacterial communities in full-scale wastewater treatment systems," *World Journal of Microbiology and Biotechnology*, vol *32*, no. 6, pp. 1–8, 2016, doi: 10.1007/s11274-016-2012-9.
- [50] N. Mao, H. Ren, J. Geng, L. Ding, and K. Xu, "Engineering application of anaerobic ammonium oxidation process in wastewater treatment," *World J. Microbiol. Biotechnol.*, vol. 0, no. 0, p. 0, 2017, doi: 10.1007/s11274-017-2313-7.
- [51] S. gang Su, H. yi Cheng, T. ting Zhu, H. cheng Wang, and A. jie Wang, "Kinetic competition between microbial anode respiration and nitrate respiration in a bioelectrochemical system," *Bioelectrochemistry*, vol. 123, pp. 241–247, 2018, doi: 10.1016/j.bioelechem.2018.06.00.
- [52] S. K. Al-Amshawee, M. Y. Yunus, and A. A. Azoddein, "A Review Study of Biofilm Bacteria and Microalgae Bioremediation for Palm Oil Mill Effluent: Possible Approach," *IOP Conf. Ser. Mater. Sci. Eng.*, vol. 736, no. 2, 2020, doi: 10.1088/1757-899X/736/2/022034.
- [53] J. Ma, Z. Wang, D. He, Y. Li, and Z. Wu, "Long-term investigation of a novel electrochemical membrane bioreactor for lowstrength municipal wastewater treatment," *Water Res.*, vol. 78, pp. 98–110, 2015, doi: 10.1016/j.watres.2015.03.033.

- [54] M. H. Gerardi, Wastewater Bacteria. 2006, doi: 10.1002/0471979910.
- [55] A. Jing, T. Liu, X. Quan, S. Chen, and Y. Zhang, "Enhanced nitrification in integrated floating fixed-film activated sludge (IFFAS) system using novel clinoptilolite composite carrier," *Front. Environ. Sci. Eng.*, 2019, doi: 10.1007/s11783-019-1153-0.
- [56] X. Zhang, X. Chen, C. Zhang, H. Wen, W. Guo, and H. Hao, "Bioresource Technology Effect of filling fraction on the performance of sponge-based moving bed biofilm reactor," *Bioresour. Technol.*, vol. 219, pp. 762–767, 2016, doi: 10.1016/j.biortech.2016.08.031.
- [57] S. K. Al-Amshawee and M. Y. M. Yunus, "A review on possible approaches of anaerobic biological processes for palm oil mill effluent: Process, quality, advantages, and limitations," *IOP Conf. Ser. Mater. Sci. Eng.*, vol. 702, no. 1, 2019, doi: 10.1088/1757-899X/702/1/012058.
- [58] Y. Wang, D. Wang, Q. Yang, G. Zeng, and X. Li, "Wastewater Opportunities for Denitrifying Anaerobic Methane Oxidation," *Trends Biotechnol.*, vol. 35, no. 9, pp. 799–802, 2017, doi: 10.1016/j.tibtech.2017.02.010.
- [59] O. Modin, K. Fukushi, and K. Yamamoto, "Denitrification with methane as external carbon source," vol. 41, pp. 2726–2738, 2007, doi: 10.1016/j.watres.2007.02.053.
- [60] I. Schmidt and E. Bock, "Anaerobic ammonia oxidation with nitrogen dioxide by Nitrosomonas eutropha," pp. 106–111, 1997, doi: 10.1007/s002030050422.
- [61] Y. V Nancharaiah and G. K. K. Reddy, "Aerobic granular sludge technology: mechanisms of granulation and biotechnological applications," *Bioresour. Technol.*, vol. 247, pp. 1128-1143, 2017, doi: 10.1016/j.biortech.2017.09.131.
- [62] T. Khin and A. P. Annachhatre, "Novel microbial nitrogen removal processes," vol. 22, pp. 519–532, 2004, doi: 10.1016/j.biotechadv.2004.04.003.
- [63] G. Carvalho, P. C. Lemos, A. Oehmen, and M. A. M. Reis, "Denitrifying phosphorus removal: Linking the process performance with the microbial community structure," vol. 41, pp. 4383–4396, 2007, doi: 10.1016/j.watres.2007.06.065.
- [64] H. T. T. Nguyen, V. Q. Le, A. A. Hansen, J. L. Nielsen, and P. H. Nielsen, "High diversity and abundance of putative polyphosphate-accumulating Tetrasphaera-related bacteria in activated sludge systems," *FEMS Microbiol. Ecol.*, vol. 76, no. 2, pp. 256–267, 2011, doi: 10.1111/j.1574-6941.2011.01049.x.
- [65] A. T. Mielczarek, H. Thi, T. Nguyen, J. L. Nielsen, and P. H. Nielsen, "Population dynamics of bacteria involved in enhanced biological phosphorus removal in Danish wastewater treatment plants," *Water Res.*, vol. 47, no. 4, pp. 1529–1544, 2012, doi: 10.1016/j.watres.2012.12.003.
- [66] P. H. Nielsen *et al.*, "A conceptual ecosystem model of microbial communities in enhanced biological phosphorus removal plants," *Water Res.*, vol. 44, no. 17, pp. 5070–5088, 2010, doi: 10.1016/j.watres.2010.07.036.
- [67] M. Terashima, A. Yama, M. Sato, I. Yumoto, Y. Kamagata, and S. Kato, "Culture-dependent and -independent identification of polyphosphate-accumulating Dechloromonas spp. Predominating in a full-scale oxidation ditch wastewater treatment plant," *Microbes Environ.*, vol. 31, no. 4, pp. 449–455, 2016, doi: 10.1264/jsme2.ME16097.
- [68] K. L. Sublette and N. D. Sylvester, "Oxidation of hydrogen sulfide by Thiobacillus denitrificans: Desulfurization of natural gas," *Biotechnol. Bioeng.*, vol. 29, no. 2, pp. 249–257, 1987, doi: 10.1002/bit.260290216.
- [69] K. L. Straub, M. Benz, B. Schink, and F. Widdel, "Anaerobic, Nitrate-Dependent Microbial Oxidation of Ferrous Iron," vol. 62, no. 4, pp. 1458–1460, 1996, doi: 10.1128/aem.62.4.1458-1460.1996.
- [70] A. K. Ghattas, F. Fischer, A. Wick, and T. A. Ternes, "Anaerobic biodegradation of (emerging) organic contaminants in the aquatic environment," *Water Res.*, vol. 116, pp. 268–295, 2017, doi: 10.1016/j.watres.2017.02.00.
- [71] J. T. Moraghan and R. J. Buresh, "Chemical Reduction of Nitrite and Nitrous Oxide by Ferrous Iron," Soil Sci. Soc. Am. J., vol. 41, no. 1, pp. 47–50, 1977, doi: 10.2136/sssaj1977.03615995004100010017x.
- [72] S. Jagadevan, M. Jayamurthy, P. Dobson, and I. P. Thompson, "A novel hybrid nano zerovalent iron initiated oxidation e Biological degradation approach for remediation of recalcitrant waste metalworking fluids," *Water Res.*, vol. 46, no. 7, pp. 2395–2404, 2012, doi: 10.1016/j.watres.2012.02.006.
- [73] Y. Yang, B. Li, S. Zou, H. H. P. Fang, and T. Zhang, "Fate of Antibiotic Resistance Genes in Sewage Treatment Plant Revealed by Metagenomic Approach," *Water Res.*, 2014, doi: 10.1016/j.watres.2014.05.019.
- [74] L. Rizzo *et al.*, "Science of the Total Environment Urban wastewater treatment plants as hotspots for antibiotic resistant bacteria and genes spread into the environment: A review," *Sci. Total Environ.*, vol. 447, pp. 345–360, 2013, doi: 10.1016/j.scitotenv.2013.01.032.
- [75] D. Li, R. Qi, M. Yang, Y. Zhang, and T. Yu, "Bacterial community characteristics under long-term antibiotic selection pressures," *Water Res.*, vol. 45, no. 18, pp. 6063–6073, 2011, doi: 10.1016/j.watres.2011.09.002.
- [76] A. Novo, S. André, P. Viana, O. C. Nunes, and C. M. Manaia, "Antibiotic resistance, Antimicrobial residues and bacterial community composition in urban wastewater," *Water Res.*, vol. 47, no. 5, pp. 1875–1887, 2013, doi: 10.1016/j.watres.2013.01.010.
- [77] K. Huang, J. Tang, X. Zhang, K. Xu, and H. Ren, "A Comprehensive Insight into Tetracycline Resistant Bacteria and Antibiotic Resistance Genes in Activated Sludge Using Next-Generation Sequencing," pp. 10083–10100, 2014, doi: 10.3390/ijms150610083.