

COMPARISON STUDY OF ESSENTIAL OIL EXTRACTED FROM *ETLINGERA ELATIOR* (TORCH GINGER) BY USING SOXLET EXTRACTION

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ABSTRACT

Significance of essential oils from *Etingera elatior* (*E. elatior*) are growing these days and have prospective to embrace great export in future, yet the quantity of the essential oil extracted does not extant in decent rate. Therefore the purpose of this research is to determine the optimum yield of the *E. elatior* essential oil. The *E. elatior* were prepared in two conditions which are normal (fresh) and dry grinded sample. The extraction of normal and dry grinded *E. elatior* was conducted using Soxhlet extraction with methanol as a solvent. The extraction process was conducted at different extraction time (2, 4, 6 hours). The study indicated that the dry grinded *E. elatior* produced a higher yield of essential oil as compared to normal grinded *E. elatior*. The yield of essential oil obtained from dry grinded *E. elatior* was 71.44% while with normal grinded *E. elatior*, the yield was 56.42%. The effect of extraction time towards yield of essential oil shown that prolong extraction up to 6 hours gave a higher yield of *E. elatior* essential oil. The constituents in the *E. elatior* essential oil were determined and quantified using GC-MS analysis. The analysis indicated that the *E. elatior* essential consist of 2-Furaldehyde, 5-methyl, 3-Methyl-1,2-cyclopentanedione, Maple lactone, Furan-2,5-dial, Pyranone and 5-Hydroxymethyl furaldehyde.

Keywords: Torch Ginger, *Etingera elatior*, Essential Oil, Soxhlet Extraction, GC-MS.

1.0 INTRODUCTION

Etingera elatior (*Zingiberaceae*) or its mutual name torch ginger and widespread acknowledged in Malaysian as “bunga kantan” is symbolize as a natural to Asia and one of the most attractive of all tropical blossoming plants. There are broad traditional used of *E. elatior* plant in Malaysia. For example, the new shoots, flower sprouts or fruits can be consumed directly by ethnic communities or as a condiment or cooked. Jeevani (2010) stated that the flower also has been used as a lawn ornamel because the inflorescences or blossom of *E. elatior* are very common as a seasoning of nutrition additive and crucial elements in some traditional foods of Malaysia like asam laksa and nasi kerabu. *E. elatior* essential oil is extensively used as natural essences in food, scents in fragrance, and in traditional medicines (Maui, 2013). Chan (2011) reported that among 5 *Etingera* species investigated, *E. elatior* has the highest antioxidant properties. Hazrin (2008) stated that the worldwide demands of *E. elatior* have opened up the worldwide opportunities due to its increasing application in pharmaceutical, aromatherapy and also cosmetics.

Extraction of essential oil from *E. elatior* can be obtained using hydro distillation or solvent extraction. However this process results in low yield, losses of its volatile compounds, residue of toxic solvent, very long extraction times and also degradation of unsaturated compound (Mostafa *et al.*, 2013). Moreover, steam distillation always involved with a higher operating cost (Silva *et al.*, 2013). Another recent technology in essential oil extraction is supercritical extraction. Unfortunately, supercritical CO₂ method is very expensive and required a higher operating of time (Barve *et al.*, 2010). Thus in order to overcome these limitations, optimization of the extraction process is needed to achieve higher extraction yield with minimum operation cost. It is reported by L.F Hu (2005), extraction is influenced by several factors including extraction time, pressure, temperature, and sample mass. Therefore, the present was focused on the optimization of the extraction process using Soxhlet extraction by varying extraction time and morphology of the raw material used in the extraction.

2.0 METHODOLOGY

Preparation of raw material (*E. elatior* inflorescence)

Two different samples (normal (fresh) and dry grinded) of *E. elatior* were investigated. In the present study, *E. elatior* inflorescences were brought from the market in Shah Alam, Selangor. The inflorescences were taken from *E. elatior*, washed with tap water to remove dirt and stain. The broken and gloomy inflorescences were detached physically and were dried for 9 hours at 40°C as according to the method describable by Hazrin (2008).

Extraction of essential oil

25g of *E. elatior* inflorescence was extracted with 200 mL of methanol by using Soxhlet extraction apparatus (Model: Mtop / Korea, Series Number of 2706) at 2, 4 and 6 hours of extraction time. The mixture was then evaporated in the rotary evaporator (Buchi V-700 Laboratory Rotary Evaporator with Vacuum Pump) for 45 minutes. The moisture content and oil yield were calculated by using the formula in Equation 1 and 2, respectively.

$$\% \text{ Humidity} = \frac{\text{Mass before dry} - \text{Mass after dry}}{\text{Mass after dry}} \times 100 \quad (1)$$

$$\text{Percent yield, \%} = \frac{\text{Mass of sample after extraction (g)}}{\text{Mass of sample before extraction (g)}} \times 100 \quad (2)$$

Gas Chromatography Mass Spectrometry (GC-MS)

The determination the concentration of chemical constituents in essential oil of *E. elatior* was detected using the GC-MS analyses which were performed via Varian 450 gas chromatograph with attached Varian 240 mass spectrometer. This apparatus was equipped 70 eV with capillary column (30 m × 0.25 mm × film thickness 0.25 μm) at intermediate polarity. Injector temperature was set at 250 °C and oven temperature was programmed initially 60 °C with 2 min hold then 60 to 120 °C (2 min hold) with 3°C min⁻¹ temperature programming and 120 to 270 °C at a rate of 2 °C min⁻¹. Injection volume was 1 μL with average velocity of 37 cm s⁻¹. Helium was used as carrier gas and the flow rate was set at 1 mL min⁻¹ (Faridahanim *et al.*, 2007).

3.0 RESULTS AND DISCUSSION

Extraction Yield at different *E. elatior* morphology

Table 1 show the moisture content of the prepared *E. elatior* before extraction was conducted. The purposed of the drying is to maximum the essential oil extracted from the material. The average of the moisture content for the dried sample was 7.13%. The dried *E. elatior* used in the extraction was controlled its moisture content to the value below than 10%. The yield of the essential oil at different morphology of *E. elatior* is shown in Figure 1.

Table 1: Moisture Content of dried *E. elatior*.

Sample	Sample Mass (g)	Final Mass (g)	Moisture Content, x (%)
A	38.5748	35.5375	8.55
B	33.8377	31.4399	7.63
C	33.4802	31.3310	6.86

The results indicated that normalgrinded *E. elatior* gave lower yield of the *E. elatior* essential oil as compared to the dried grinded *E. elatior*. It shows that the dried grinded *E. elatior* gave 21 – 30% higher of extraction yield.

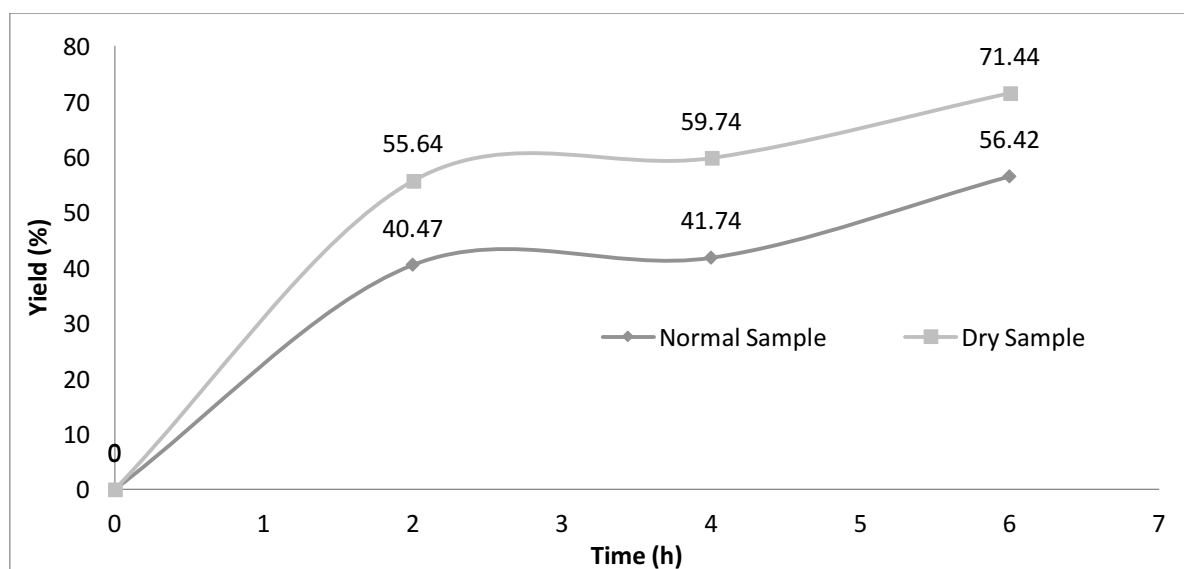


Figure 1: Percent Yield of *E. elatior* Essential Oil

While for the effect of extraction time on the yield of the essential oil indicated that prolong of the extraction period capable to increase the yield of the essential oil for normal and dry grinded of *E. elatior*. The results show that 6 hours extraction time produced the highest essential oil up to 56.42 % and 71.44 % oil yield for normal and dried grinded *E. elatior* respectively. Both samples showed an increment of essential oil yield as the extraction time increased and dry *E. elatior* is preferable to enhance yield of extraction. The result is in agreement with the finding by Jeevani *et al.*, (2011) which determine that drying might reduce the amount of oil in each plant, but can greatly increase the yield per batch. According to Bence (2008) moisture contents between 7 and 18% had little effect on extractability of oil. However, the water had a negative entrainer

effect by extraction of samples containing more than 18% water. When the moisture content was increased, a decrease in efficiency was found. In the case of high moisture content (~85%) the extraction was almost inhibited.

Chemical Constituent of *E. elatior* Essential Oil

The extracted oil was analyzed for its chemical constituent via a GC-MS. Each component in the extracted oil was identified through matching with data of various components in the mass spectrometer library. The GCMS spectrum of *E. elatior* is shown in Figure 2 and Table 2 listed the outcome of the matching process, indicating the quality of the matching in terms of percentage, following the international standard compounds of *E. elatior* essential oil.

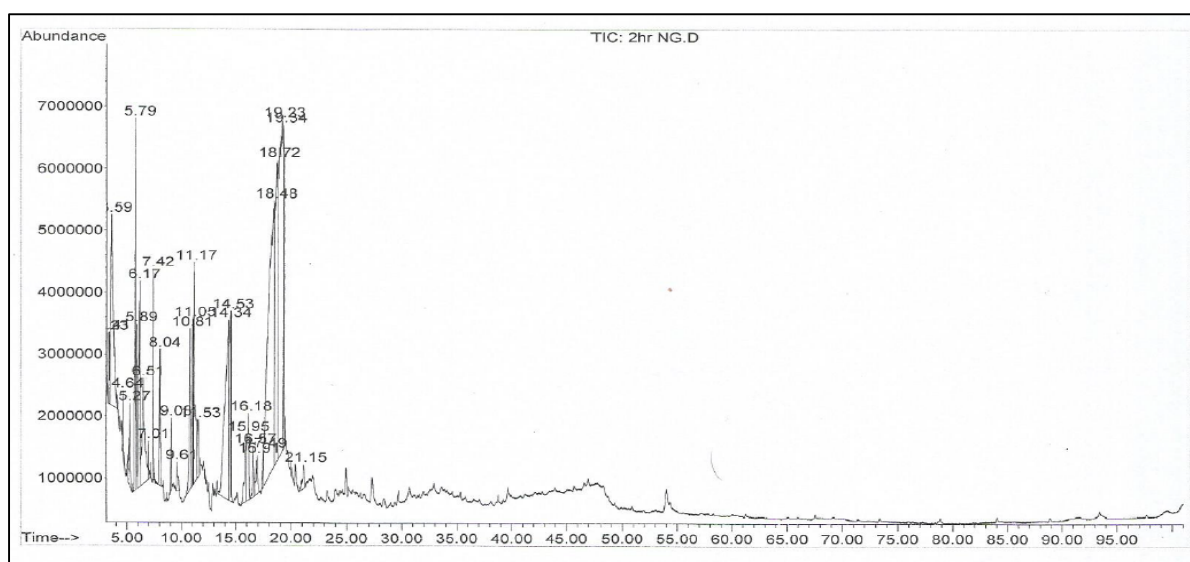


Figure 2: The Chemical constituent of the *E. elatior* Essential Oil for 2 Hours Extraction of Normal Sample

Table 2: Constituent of *E. elatior* Essential Oil Natural Samples (2 Hours Extraction)

RT (min)	Component	CAS #	Qual (%)
3.23	(S)-[(S)-2-Phenyl-1,3,2-dioxaborolan-4-yl]-1,2-ethanediol	074807-80-0	36
3.41	2-furaldehyde	000098-01-1	35
3.59	2-furaldehyde	000098-01-1	72
4.64	Levogluosenone	037112-31-5	64
5.26	Butanoic acid, 2-ethyl-3-oxo-, methyl ester	051756-08-2	50
5.79	Oxime-, methoxy-phenyl	1000222-86-6	90
5.89	2(5H)-Furanone	000497-23-4	59
6.17	1,2-Cyclopentanedione	003008-40-0	72
6.51	2-Furancarboxaldehyde, 5-methyl	000620-02-0	53
7.01	2-Furancarboxaldehyde, 5-methyl	000620-02-0	72
7.42	3,4-Diflouronisole	115144-40-6	50
8.04	2H-Pyran-2,6(3H)-dione	005926-95-4	72

9.08	Maple lactone	000080-71-7	87
9.61	1H-Indole, 1-methyl-2-phenyl	003558-24-5	25
10.81	Furan-2,5-dial	000823-82-5	59
11.05	2-Furancarboxylic acid, hydrazide	003326-71-4	46
11.17	2,4,5-Trihydroxypyrimidine	000496-76-4	53
11.53	1,3-Bis(trimethylsilyl)benzene	002060-89-1	72
14.34	Pyranone	028564-83-2	38
14.53	Pyranone	028564-83-2	72
15.95	4H-Pyran-4-one, 3,5-dihydroxy-2-methyl	001073-96-7	55
16.18	N-Benzyloxycarbonyl-L-alanyl-L-alanine ethylamide	066382-92-1	43
16.57	1,2-Benzenediol	000120-80-9	87
16.91	5-Acetoxymethyl-2-furaldehyde	010551-53-3	53
17.49	5-(p-Aminopenyl)-4-(p-tolyl)-2-thiazolamine	094460-46-5	59
18.48	2-Furancarboxaldehyde, 5-(hydroxymethyl)	000067-47-0	91
18.72	2-Furancarboxaldehyde, 5-(hydroxymethyl)	000067-47-0	91
19.23	2-Furancarboxaldehyde, 5-(hydroxymethyl)	000067-47-0	91

Considering a minimum of 60% matches with the standard components in the library, only 7 chemicals constituent were identified from the list of chemicals constituents distinguished in all samples presents in Table 3. These components were 2-Furaldehyde, 2-Furaldehyde, 5-methyl, 3-Methyl-1,2-cyclopentanedione, Maple lactone, Furan-2,5-dial, Pyranone and 5-Hydroxymethyl furaldehyde. These chemicals represent the quality of the extracted *E. elatior* oil, which gave essential oil characteristics.

Table 3: Major Components of All *E. elatior* Essential Oil Samples

Component	CAS no.	Formula	Yield (%)					
			Normal		Dry			
			2hr	4hr	6hr	2hr	4hr	6hr
2-Furaldehyde	00009 8-01-1	C ₅ H ₄ O ₂	6.19	4.22	3.86	6.86	9.43	10.42
2-Furaldehyde, 5-methyl	00062 0-02-0	C ₆ H ₆ O ₂	4.72	3.92	2.22	4.64	5.53	5.56
3-Methyl-1,2-cyclopentanedione	00076 5-70-8	C ₆ H ₈ O ₂	0.52	0.63	0.97	0.42	0.46	0.56
Maple lactone	00008 0-71-7	C ₆ H ₁₀ O ₃	0.52	0.63	0.97	0.42	0.46	0.56
Furan-2,5-dial	00082 3-82-5	C ₆ H ₄ O ₃	2.78	3.21	3.36	2.78	2.74	2.3
Pyranone	02856 4-83-2	C ₆ H ₈ O ₄	11.87	11.94	12.47	10.82	10.78	10.55
5-Hydroxymethyl furaldehyde	00006 7-47-0	C ₆ H ₆ O ₃	50.41	49.59	45.67	52.36	44.65	39.61
Others			22.99	25.86	30.48	21.7	25.95	30.44
Total			100	100	100	100	100	100

All of the constituent present in the *E. elatior* has various industrial applications. For example, *5-Hydroxymethyl furaldehyde* is important as flavoring representatives in foods (Li *et. al.*, 1988) and showed the biggest peak range tailed by *Pyranone*, a natural ingredients and extractives. *2-Furaldehyde* gave the scent of almond while *2-Furaldehyde*, *5-methyl* are skillful of foraging peroxyinitrite. *3-Methyl-1,2-cyclopentanedione* is used as cockroach attractant as reported by Blanco *et. al.*(1991). Moreover, *Maple lactone* is function as a regular constituents and extractives and *2 Furan-2,5-dial* is one of the flavoring agent in numerous food elements (Meidell and Filipello,1969). Besides, *5-Hydroxymethyl furaldehyde* is used in the production of dialdehydes, glycols, ethers, aminoalcohols, acetals and in aqueous acid to catalyses ring opening.

4.0 CONCLUSIONS

The extraction of *E. elatior* essential oil using dissimilar conditions (dry and normal) gave significant impact on the percentage of essential oil yield. Dry grinded *E. elatior* produced 21-30 % higher yield as compared to normal *E. elatior*. While, prolong of the extraction time up to 6 hours result up to 71.4% of the essential oil yield. The chemical composition of essential oil after extracted in Soxhlet extraction method determined by GC-MS quantified 7 chemical components that exist the essential oil which are *2-Furaldehyde*, *5-methyl*, *3-Methyl-1,2-cyclopentanedione*, *Maple lactone*, *Furan-2,5-dial*, *Pyranone* and *5-Hydroxymethyl furaldehyde*. These chemical constituents have a great potential in various industrial application.

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REFERENCES

- Barve K. H., Laddha, K. S., & Jayakumar, B. (2010). Extraction of saponins from Safed Musli. *Pharmacognosy Journal*, 2: 561–564.
- Bence N., B. Simandi. 2008. Effects Of Particle Size Distribution, Moisture Content, And Initial Oil Content On The Supercritical Fluid Extraction Of Paprika. *The Journal of Supercritical Fluids*. Volume 46, (Issue 3): 293–298.
- Blanco Gomis D., Gutiérrez Alvarez, M.D. Sopena Naredo, L. & Mangas Alonso, J.J.(1991) High-Performance liquid chromatographic determination of furfural and hydroxymethylfurfural in apple juices and concentrates. *Chromatographia*, 32:45-48.
- Chan E. W.C., Lim Y.Y. & Wong S. K. (2011). Phytochemistry and Pharmacological Properties. Of *Etingera elatior*: A Review, UCSI University, Faculty of Applied Sciences Kuala Lumpur, Malaysia.
- Faridahanim M. J., Che P. O., & Nor H. I., Khalijah A. (2007). Analysis of Essential Oils of Leaves, Stems, Flowers Andrhizomes of *Etingera Elatior* (Jack) R. M. Smith. *The Malaysian Journal of Analytical Sciences*, 11(1): 269-273.
- Han C. V., Bhat R. & Rusul G. (2012). Flower Extracts and Their Essential Oils as Potential Antimicrobial Agents for Food Uses and Pharmaceutical Applications. *Comprehensive Reviews in Food Science and Food Safety*, 11(1): 34–55.

- Hazrin A. (2008). Monitoring of Quality of Essential Oil from *Etlingera* sp. 2 (Zingiberaceae) by GC and GC-MS. Universiti Malaysia Pahang: Faculty of Chemical and Natural Resources Engineering.
- Jeevani M. M. O. W., Bhat R. & Alias A. K. (2011). Effect of Extraction Solvents on the Phenolic Compounds and Antioxidant Activities of Bunga Kantan (*Etlingera elatior* Jack.) Inflorescence. *Journal of Food Composition and Analysis*, 24: 615-619.
- Jeevani O. W., M. M., Karim, A. A. & Bha R. (2010). Evaluation of nutritional quality of torch ginger (*Etlingera elatior* Jack.) inflorescence. *International Food Research Journal*, 18(4): 1415-1420.
- L.F Hu, S.P.Li, H.Cao, J.J.Liu, J.L.Gao F.Q.Yang, Y.T Wang. 2005. GC-MS Fingerprint of *Pogostemon Cablin* in China. Institute of Chinese Medical Science, University of Macau.
- Li Z-F., Sawamura, M. & Kusunose, H. (1988) Rapid determination of furfural and 5-hydroxymethylfurfural in processed citrus juices by HPLC. *Agric. Biol. Chem.*, 52: 2231–2234.
- Maui M. (11 April, 2013). Hawaiian Ginger:Not Just For Eating. Retrieved on 6 April, 2014, from The Private Naturalist <https://theprivatenaturalist.wordpress.com/tag/torch-ginger>.
- Meidell E. & Filipello, F. (1969) Quantitative Determination of Hydroxymethylfurfural in Sherries and Grape Concentrate. *Am. J. Enol. Viticult*, 20(3):164-168
- Mostafa A., Sudisha, J., El-Sayed, M., Ito, S. -I., Yamauchi, N., & Shigyo, M. (2013) Aginoside saponin, a potent antifungal compound, and secondary metabolite analyses from *Allium nigrum* L. *Phytochemistry Letters*, 6: 274–280.
- Silva L. P., Pereira, M. J., Azevedo, J., Gonçalves, R. F., Valentão, P., de Pinho P. G., & Andradea P. B. (2013) *Glycine max* (L.) Merr., *Vigna radiate* L. and *Medicago sativa* L. sprouts: A natural source of bioactive compounds. *Food Research International*, 50:167–175.